# Review

# The Use of CO<sub>2</sub> Laser in the Treatment of Peri-implantitis

George Romanos,<sup>1,2</sup> Hua-Hsin Ko,<sup>3</sup> Stuart Froum,<sup>3</sup> and Dennis Tarnow<sup>3</sup>

# Abstract

Different techniques have been used for the treatment of peri-implant defects. However, there are always questions about the issue of reosseointegration. The present paper explores the recent literature on the topic of peri-implantitis therapy, and presents a surgical protocol for implant surface decontamination using the  $CO_2$  laser, grafting of the defect, and coverage with a membrane according to a clinical case. The results appear to be promising and may improve the long-term clinical outcomes of failing dental implants.

# Introduction

**E**NDOSSEOUS IMPLANTS have a high survival rate in the restoration of fully or partially edentulous patients.<sup>1</sup> However, over a 5-y period, up to 14.4% of dental implants demonstrated peri-implant inflammatory reactions associated with bone loss.<sup>2</sup>

Implant failure has classically been attributed to bacterial infection, surgical trauma, premature fixture overload, faulty or incorrect prosthetic design, and/or improper surgical placement. The etiology of failure is thought to be infection, if there is bleeding, suppuration, pain, and high plaque and gingival indices (Figs. 1 and 2). This infectious process with progressive bone loss (Fig. 3) seen over time is categorized as peri-implantitis.<sup>3</sup>

Ideally, bone-to-implant contact should be increased and implants should become reosseointegrated. At present, there is no evidence about the utility of anti-infective treatment to prolong the longevity of an implant. There is also insufficient evidence to support any specific treatment strategy with respect to treatment of peri-implantitis.<sup>4,5</sup>

Numerous treatments have been recommended for peri-implantitis. Guided bone regeneration (GBR) has been used for the treatment of peri-implant bony defects;<sup>6–8</sup> however, this procedure has limited efficacy.<sup>9</sup> Currently, there are no clinical studies or case series documenting successful regenerative procedures in peri-implant bony lesions. Some case series demonstrated limited bone filling after GBR procedures.<sup>6</sup> To enhance the results, some investigators have suggested that it is necessary to decontaminate defective implant surfaces.<sup>6,10–13</sup> Several different methods of implant decontamination have been proposed.<sup>13</sup> Neither subgingival irrigation with local disinfectants<sup>12,14,15</sup> nor local antibiotic therapy with tetracycline fibers provided conclusive therapeutic effects.<sup>16</sup> Systemic administration of antibiotics has also been used in the treatment of peri-implantitis; however, the success was limited due to resistant strains of bacteria and ineffective drug dosages.<sup>17,18</sup> Citric acid application with sandblasting,<sup>7,19</sup> sandblasting alone,<sup>20–22</sup> or chlorhexidine irrigation<sup>23</sup> have also been recommended. However, implant decontamination using sandblasting units has been associated with risks such as emphysema.<sup>24</sup>

In contrast, encouraging results in dogs have been reported using the  $CO_2$  laser for decontamination to improve reosseointegration.<sup>25</sup> Findings of this animal study suggested that the laser may be an effective therapeutic modality in the treatment of peri-implantitis.

The purpose of the present paper was to review the literature on the use of the  $CO_2$  laser in the treatment of peri-implantitis, and to discuss the indications, advantages, and disadvantages of this technique.

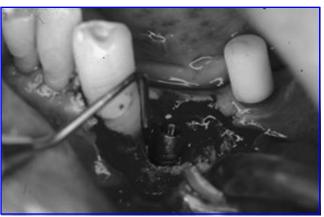
#### Materials and Methods

The literature search using MEDLINE at the Waldman Library of the New York University College of Dentistry Kriser Dental Center included a review of 71 articles from peer-reviewed journals published in English from January 1986 to December 2007. The keywords utilized were "carbon dioxide laser" and "implant" (36 articles); "laser" and "peri-implantitis" (23 articles); and "CO<sub>2</sub> laser decontamination" (12

<sup>&</sup>lt;sup>1</sup>Divisions of Periodontology and General Dentistry, Unit of Laser Dentistry, Eastman Dental Center, University of Rochester, Rochester, New York, <sup>2</sup>Department of Oral Surgery and Implant Dentistry, Dental School (Carolinum), University of Frankfurt, Frankfurt, Germany, and <sup>3</sup>Department of Periodontology and Implant Dentistry, College of Dentistry, New York University, New York, New York.



**FIG. 1.** Deep peri-implant pockets associated with tissue bleeding.



**FIG. 3.** Measurement of a peri-implant pocket with a periodontal probe (7 mm).

articles). In addition, this paper presents the surgical protocol of the treatment of peri-implantitis using a  $CO_2$  laser according to a clinical case report.

#### Results

The results of this literature review are presented in Table 1. The data include the type of study, decontamination type, implant surface, augmentation procedure (bone grafting material), and the results. The literature analysis is focused on the use of the  $CO_2$  laser for implant surface decontamination and the treatment of peri-implantitis.

Most of the *in vitro* studies using the CO<sub>2</sub> laser showed no structural changes in the implant surface. In most studies a significant degree of bacterial reduction has been documented. *In vivo* animal studies demonstrated promising results with reosseointegration occurring after CO<sub>2</sub> laser decontamination. In one human clinical study,<sup>37</sup> sufficient decontamination led to new bone formation. According to this experience we used a similar protocol to decontaminate the surface of defective implants. After elevation of a mucoperiosteal flap, removal of granulation tissue with plastic curettes, decontamination with a CO<sub>2</sub> laser (Figs. 4, 5, and 6; continuous mode, power levels of 2–4 W, with a non-



**FIG. 2.** Peri-implant bony defect due to bacteria accumulation (peri-implantitis).

contact defocused handpiece), augmentation with bone grafting material, use of an absorbable membrane (GBR), and flap closure, we found good healing and new bone formation compared to baseline (Fig. 7). Recent clinical studies using the  $CO_2$  laser to decontaminate implant surfaces and bone filling with autogenous bone or bone grafting materials showed good results and long-term success of the oral implants.<sup>38</sup>

# Discussion

Non-surgical methods to treat peri-implantitis include mechanical instrumentation and the use of a variety of antibacterial agents. The antibiotic treatment of peri-implantitis may not allow sufficient bone filling or reosseointegration in deep peri-implant bony defects. Surgical therapy may be necessary to treat peri-implant defects,<sup>10,11,13</sup> however, to date no treatment method has attained consistent long-term results.

With the protocol described above it is possible to efficiently decontaminate the implant surface and to augment the peri-implant bony defects with either autogenous bone or bone replacement graft materials. In an animal model, osseous filling and reosseointegration were observed. This conclusion was supported by the histological observations by Deppe et al.<sup>25</sup> and Stübinger et al.,<sup>34</sup> who noted that reosseointegration occurred and bone filling was induced in peri-implant defects after CO<sub>2</sub> laser irradiation.

The physical properties of the laser energy and its interaction with tissues, due to reflection, scattering, transmission, and absorption, may explain why the implant surface can be decontaminated in all areas, including within the threads. The light may induce these antibacterial effects due to its absorption by the implant and the surrounding tissues, or it may be reflected by the metal surface, causing a slight elevation in tissue temperature.

The temperature changes seen during  $CO_2$  laser irradiation have been studied extensively.<sup>29,39–41</sup> Linear increases to temperatures >50°C were observed with increases in power levels and exposure times, and the pulse mode generated significantly less heat. The results of this study suggest that caution should be exercised when using the  $CO_2$  laser for second-stage dental implant surgery, as the temperatures of

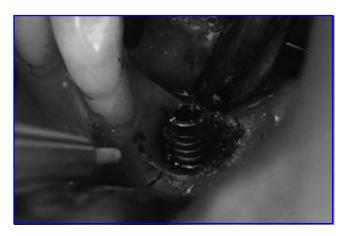
| Author                            | Year | Study type                           | Decontamination<br>technique                         | Implant<br>surface              | Guided bone<br>regeneration              | Results  |
|-----------------------------------|------|--------------------------------------|--|---------------------------------|--|--|
| $Ganz^{26}$                       | 1994 | In vitro study (pig                  | CO <sub>2</sub>                                      | Hydroxyapatite                  |  | Minimal temperature change                             |
| Oyster et al. <sup>27</sup>       | 1995 | In vitro study (pig                  | CO <sub>2</sub>                                      | coalect                         |  | Time and power affected the                            |
| Kato et al. <sup>28</sup>         | 1998 | In vitro study<br>(titanium discs)   | CO <sub>2</sub><br>CO <sub>2</sub>                   | Machined,<br>sandblasted        |  | Efficiently decontaminate<br>bacteria without altering |
| Mouhyi et al. <sup>29</sup>       | 1999 | In vitro study<br>(titanium foil)    | CO <sub>2</sub> + hydrogen<br>peroxide + citric acid | TiO <sub>2</sub>                |  | Re-establishment of the atomic composition and         |
| Romanos et al. <sup>30</sup>      | 2001 | In vitro (titanium                   | CO <sub>2</sub>                                      | Various                         |  | vaue suucine<br>No surface modifications               |
| Deppe et al. <sup>25</sup>        | 2001 | In vivo study                        | $CO_2$ , $CO_2$ + sandblasting                       | SdL                             | e-PTFE only                              | New bone formation                                     |
| Romanos et al. <sup>31</sup>      | 2002 | In vitro (titanium                   | CO <sub>2</sub>                                      | Sandblasted                     |  | Bacterial reduction                                    |
| Kreisler et al. <sup>32</sup>     | 2002 | uiscs)<br>I <i>n vitro</i> (titanium | CO <sub>2</sub>                                      | Various                         |  | No damage  |
| Persson et al. <sup>33</sup>      | 2004 | In vivo study<br>(beagle dogs)       | CO <sub>2</sub> + hydrogen<br>peroxide + systemic    | Sandblasted<br>large-grit acid- | ×  | New bone formation                                     |
| Stübinger et al. <sup>34</sup>    | 2005 | In vivo study                        | antibiotics<br>CO <sub>2</sub>                       | ercnea, macrunea<br>TPS         | ×  | New bone formation                                     |
| Park et al. <sup>35</sup>         | 2005 | (beagle dogs)<br>In vitro (titanium  | CO <sub>2</sub>                                      | Various                         |  | No damage  |
| Romanos et al. <sup>36</sup>      | 2006 | uiscs)<br>In vitro (titanium         | CO <sub>2</sub>                                      | Various                         |  | Osteoblast growth and                                  |
| Deppe et al. <sup>37</sup>        | 2007 | In vivo study                        | CO <sub>2</sub>                                      | Various                         | $\beta$ -TCP + e-<br>DTFF                | Autocompetition  |
| Romanos and Nentwig <sup>38</sup> | 2008 | In vivo study<br>(human)             | CO <sub>2</sub>                                      | Various                         | Autogenous<br>bone or<br>BioOss +<br>GBR | New bone formation                                     |

Table 1. Studies of  $\mbox{CO}_2$  Laser Interaction with Dental Implants

e-PTFE, expanded polytetrafluoroethylene; β-TCP, β-tricalcium phosphate; TPS, titanium plasma sprayed.



FIG. 4. Handpieces for CO<sub>2</sub> laser irradiation.



**FIG. 5.** Decontamination of the implant surface and the surrounding tissues with a  $CO_2$  laser (4 W, continuous mode, with a non-contact defocused handpiece).

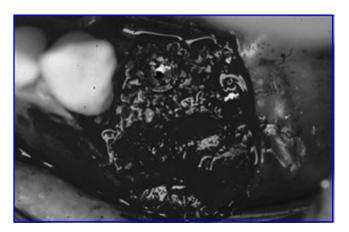


FIG. 6. Augmentation with BioOss® (Shirley, NY).



**FIG. 7.** Radiographic examination 3 y after loading showing excellent formation of new calcified (bone-like) tissue.

dry surfaces exceeded the accepted thresholds for bone damage at clinically relevant settings. However, these temperature changes were measured at the implant-bone interface. Studies have shown that the CO<sub>2</sub> laser produced minimal temperature changes in continuous mode at power levels <4 W, and when used in pulsed mode. Finally, Kreisler et al.<sup>41</sup> concluded that implant surface decontamination with both laser types (CO<sub>2</sub> and GaAlAs) must be time-limited to allow the implant and bone to cool down.

# Conclusion

The use of the  $CO_2$  laser in the treatment of periimplantitis deserves consideration as an efficacious treatment modality, as there appears to be little risk to the patient. However, the surgeon requires special training with respect to safety procedures and laser-tissue interactions. In addition, the cost of the laser unit and its wavelength must also be taken into consideration.

Further clinical and histological research is necessary to determine if long-term success can be achieved, and if reosseointegration affects implant survival and controls the disease process. Special studies using systems with different implant designs and various implant surfaces may be useful to assess the efficacy of this treatment modality.

#### **Disclosure Statement**

No conflicting financial interests exist.

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Address reprint requests to: Dr. Georgios Romanos Eastman Dental Center Unit of Laser Dentistry University of Rochester 625 Elmwood Avenue Rochester, New York 14620

E-mail: Georgios\_Romanos@urmc.rochester.edu